

Two cannabidiol metabolites formed by rat liver

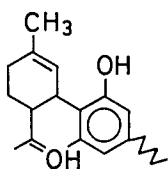
Cannabis sativa L. contains three major cannabinoids, *i.e.* Δ^1 -tetrahydrocannabinol (Δ^1 -THC*), cannabinol (CBN) and cannabidiol (CBD; I). It appears that only Δ^1 -THC is psychotomimetically active but its effect is apparently modified by other compounds also present in Cannabis (*cf.* Mechoulam, 1970). From recent evidence (Paton & Pertwee, 1972; Jones & Pertwee, 1972) it is likely that CBD contributes to the activity of cannabis preparations by inhibiting the metabolism both of Δ^1 -THC and its primary active metabolite 7-hydroxy- Δ^1 -THC.†

We have now studied the metabolism of CBD using rat liver homogenate 10 000 g supernatant.

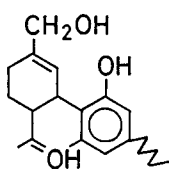
[14 C] CBD was prepared from [14 C] olivetol (Nilsson, Nilsson & Agurell, 1969) using the method of Petrzilka, Haefliger & Sikemeier (1969). The labelled CBD (30 mg) was incubated with 10 000 g supernatant from rat liver homogenate using an *in vitro* system described by Tagg, Yasuda & others (1967) with the modifications previously reported (Nilsson, Agurell & others, 1970).

The incubation mixture was extracted with light petroleum to remove most of the unchanged CBD. Since one possible metabolic transformation of CBD could be a cyclization to Δ^1 -THC, the extract was analysed for Δ^1 -THC by thin-layer chromatography (Korte & Sieper, 1960) and gas chromatography. Any formed Δ^1 -THC could not be detected.

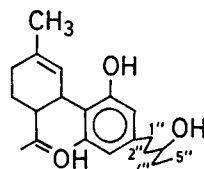
Further extractions of the incubate with diethyl ether recovered several metabolites (35% conversion of CBD) which were partially separated by column chromatography on Florisil (*cf.* Nilsson & others, 1970). One fraction of similar polarity to 7-hydroxy- Δ^1 -THC was further purified by t.l.c. on Silica gel G plates with ether-light petroleum (5:1) as solvent. The isolated material (2 mg) was found by g.l.c. to contain, apart from minor constituents, two metabolites (II and III) in the ratio 8:2.



I



II



III

The mass spectrum of the main metabolite (II) showed prominent peaks at *m/e* (rel. intensity): 330 (7, M⁺), 315 (3), 312 (35), 299 (37), 244 (100), and 231 (51). The peak at *m/e* 299 is formed by the loss of -CH₂OH. This is also a typical feature in the spectrum of 7-hydroxy- Δ^1 -THC (Nilsson & others, 1970) and indicates a hydroxylation of CBD at C-7 or possibly C-10. The nmr spectrum of the metabolite mixture, containing 80% of II, showed a singlet at δ 1.68 due to the presence of the C-10 methyl group (in CBD at δ 1.67 ppm) with only a minor peak at δ 1.82 indicative of the C-7 methyl group in III. A signal at δ 4.02 (singlet), roughly equivalent to two protons, is assigned to the -CH₂O- group of II (at δ 4.02 in 7-hydroxy- Δ^1 -THC). On the basis of this evidence the main metabolite of CBD (II) is assigned the structure 7-hydroxy-CBD. The C-7 hydroxylation of CBD is analogous to the primary C-7 hydroxylation of Δ^1 -THC and CBN (*cf.* Agurell, Dahmén & others, 1972).

* Designated as Δ^9 -THC using the benzopyran numbering system.

† 11-Hydroxy- Δ^9 -THC.

The mass spectrum of metabolite III showed prominent peaks at m/e (rel. intensity): 330 (9, M^+), 315 (4), 297 (3), 262 (26), 247 (100), and 245 (6). The presence of the fragment m/e 247 (base peak) instead of m/e 231 as in the spectrum of CBD (Budzikiewicz, Alpin & others, 1965) implied that a hydroxyl group was located in the pentyl side-chain. 1''-Hydroxy-CBD, 2''-hydroxy-CBD and 3''-hydroxy-CBD were synthesized and compared with compound III by g.l.c. t.l.c. and mass spectrometry. The syntheses of 1''-hydroxy-CBD and 2''-hydroxy-CBD are discussed in a recent review (Aguirell & others, 1972). 3''-Hydroxy-CBD was synthesized by the reaction of 1-(3,5-dihydroxyphenyl)pentan-3-one ethylene dithioketal (Fahrenholtz, 1972) and (+)-*trans*-*p*-mentha-2,8-dien-1-ol in the presence of *NN*-dimethyl-formamide dioneopentyl acetal. The resulting 3''-ethylene dithioketal derivative of CBD was treated with mercuric chloride and cadmium carbonate, followed by sodium borohydride to yield 3''-hydroxy-CBD.

G.l.c. (3% SE-30/Gas-Chrom Q, 200°) gave a retention time for 1''-hydroxy-CBD of 6.45 min, for 2''-hydroxy-CBD 6.20 min, for 3''-hydroxy-CBD 6.70 min and for III 6.70 min. By t.l.c. (alumina plates eluted with $CHCl_3$ and silica gel plates eluted with ether: light petroleum 1:1) III had the same R_F -value as 3''-hydroxy-CBD. Also, the mass spectrum of III was in agreement with that of the synthetic 3''-hydroxy-CBD. Thus, 3''-hydroxy-CBD is indistinguishable from metabolite III by t.l.c., g.l.c. and mass spectrometry.

In this connection it may be noted that $\Delta^{1(6)}$ -THC hydroxylated in the side chain at the 1''- and 3''-position, respectively, have been isolated from *in vitro* experiments with $\Delta^{1(6)}$ -THC using dog liver (Maynard, Gurney & others, 1971).

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REFERENCES

- AGURELL, S., DAHMÉN, J., GUSTAFSSON, B., JOHANSON, U.-B., LEANDER, K., NILSSON, I. M., NILSSON, J. L. G., NORDQVIST, M., RAMSAY, C. H., RYRFFELDT, A., SANDBERG, F. & WIDMAN, M. (1972). In: *Cannabis and its derivatives*. Editors: J. Crown & W. D. M. Paton. Oxford: O.U.P.
- BUDZIKIEWICZ, H., ALPIN, R. T., LIGHTNER, D. A., DJERASSI, C., MECHOULAM, R. & GAONI, Y. (1965). *Tetrahedron*, **21**, 1881-1888.
- FAHRENHOLTZ, K. (1972). *J. org. Chem.*, **37**, 2204-2205.
- JONES, G. & PERTWEE, R. C. (1972). *Br. J. Pharmac.*, **45**, 375-377.
- KORTE, F. & SIEPER, H. (1960). *Ann.*, **640**, 71-83.
- MAYNARD, D. E., GURNEY, O., PITCHER, R. C. & KIERSTEAD, R. W. (1971). *Experientia*, **27**, 1154-1155.
- MECHOULAM, R. (1970). *Science*, **168**, 1159-1166.
- NILSSON, I. M., AGURELL, S., NILSSON, J. L. G., OHLSSON, A., SANDBERG, F. & WAHLQVIST, M. (1970). *Ibid.*, **168**, 1228-1229.
- NILSSON, J. L. G., NILSSON, I. M. & AGURELL, S. (1969). *Acta. chem. scand.*, **23**, 2209-2211.
- PATON, W. D. M., PERTWEE, R. C. (1972). *Br. J. Pharmac.*, **44**, 250-261.
- PETRZILKA, T., HAEFLIGER, W. & SIEKEMEIER, C. (1969). *Helv. chim. Acta*, **52**, 1102-1134.
- TAGG, J., YASUDA, D. M., TANABE, M. & MITOMA, C. (1967). *Biochem. Pharmac.*, **16**, 143-153.